



ORIGINAL RESEARCH ARTICLE

OPEN ACCESS

ANTIBACTERIAL EFFECT OF AQUEOUS EXTRACT OF ALOE VERA AGAINST THE GRAM-POSITIVE BACTERIA STAPHYLOCOCCUS AUREUS IN MEDANI CITY - GEZIRA STATE – SUDAN- 2017

¹Yasir Hakim, ²Dalia Hamza, ³A.KH Khalil, ⁴Faiez Yousif, ⁴Abubaker Elsiddig Talha, ⁵Abdalla Khalid and ⁶Mustaque Ali

- ¹Assistant Professor of Pathology with MD Pathology, Head Unit of Microbiology, Department of Basic Medical Science, College of Medicine, Dar Al Uloom University, Riyadh, Kingdom of Saudi Arabia
²Plant Pathology Center, University of Gezira, Sudan
³Head Unit of Biochemistry, Department of Basic medical science, College of Medicine, Dar Al Uloom University, Riyadh, Kingdom of Saudi Arabia
⁴Anatomy and Histology Unit, Department of Basic medical science, College of Medicine, Dar Al Uloom University, Riyadh, Kingdom of Saudi Arabia.
⁵Microbiology Unit, Department of Basic medical science, College of Medicine, Dar Al Uloom University, Riyadh, Kingdom of Saudi Arabia
⁶Community Unit, Department of Basic medical science, College of Medicine, Dar Al Uloom University, Riyadh, Kingdom of Saudi Arabia

ARTICLE INFO

Article History:

Received 16th November, 2018
Received in revised form
14th December, 2018
Accepted 20th January, 2019
Published online 27th February, 2019

Key Words:

Aloe Vera, Antimicrobial activities, Gentamycin, Staphylococcus aureus.

ABSTRACT

The plant Aloe vera was used historically as a topical to heal wounds, various skin conditions and orally as a laxative. The Gram-positive bacterium Staphylococcus aureus is considered to be the most pathogenic species of the genus Staphylococcus, being implicated in both community-acquired and nosocomial infections. The present investigation was undertaken at the University of Gezira, Center of Plant Pathology, during the year 2015. The aim of the study was to investigate the effect of Aloe vera on Staphylococcus aureus as antibacterial activity of aqueous and alcoholic extracts of Aloe vera on inhibiting the growth of the Staphylococcus aureus against a known Antibiotics (Gentamycin) as appositve control. Three concentrations of aqueous extract of Aloe vera and the Gentamycin , (25, 50 and 100%) were tested. The aqueous suspensions of the dried Aloe vera extracts were screened for their anti-Staphylococcus aureus activity using the agar-disc diffusion method. The results obtained indicated that the highest concentration of Aloe vera aqueous extract (100%) used in this study had the highest inhibitory effects (18.2 mm) against the tested bacterium, while the other two concentrations (50 % and 25%) showed an inhibition zones of 15 mm and 12 mm, respectively. For the positive control (Gentamycin), the highest inhibition zone 16.5 mm was obtained with the higher concentration (100 %). The other concentrations (25 and 50 % showed inhibition zones of 7.75 and 6 mm, respectively. The study recommended that, further research should be done to clearly identify the active ingredients of Aloe vera and their other antimicrobial activities.

Copyright © 2019, Yasir Hakim et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Citation: Yasir Hakim, Dalia Hamza, A.KH Khalil, Faiez Yousif, Abubaker Elsiddig Talha, Abdalla Khalid and Mustaque Ali. 2019. "Antibacterial effect of aqueous extract of aloe vera against the gram-positive bacteria staphylococcus aureus in medani city - Gezira State – Sudan- 2017", *International Journal of Development Research*, 09, (02), 25555-25561.

INTRODUCTION

Aloe vera belongs to the Liliaceae family, of which there are about 360 species. It is a cactus-like plant that grows readily in hot, dry climates and currently, because of demand, is cultivated in large quantities.

*Corresponding author: Yasir Hakim

Assistant Professor of Pathology with MD Pathology, Head Unit of Microbiology, Department of Basic Medical Science, College of Medicine, Dar Al Uloom University, Riyadh, Kingdom of Saudi Arabia

The gel of *A. vera* was used to treat stomach ailments, gastrointestinal problems, skin disease, constipation, radiation injury, inflammatory effect, healing wounds and burns, ulcer and diabetes. *A. vera* products are mainly for cosmetic, pharmaceutical, nutraceuticals and food industries. The gel stimulates cell growth and enhances the restoration of damaged skin. It moisturizes the skin because it has a water holding capacity. As a drink, it protects the mucous membrane of the stomach especially when irritated or damaged. *A. vera* juice is considered helpful for relieving many types of gastrointestinal irritation and juice products are widely

available (Anderson and Phillipson., 1996). *Staphylococcus aureus* poses an important problem in hospitals, nursing homes, and other health care settings. Serious infections due to these organisms currently necessitate the use of non- β -lactam antibacterial therapy (Hackbarth and Chambers, 1989). Many hospital acquired MRSA strains are only susceptible to vancomycin (Fitzgerald *et al.*, 2001). Thus, there are strong concerns about the possible development and spread of vancomycin resistance in MRSA. Some vancomycin-resistant MRSA strains have been reported since 1996 (El-Jakee *et al.*, 2014; ALian *et al.*, 2012). Some necrosis poisons cases occur by strong acids as H_2SO_4 , it affects skin created necrosis or burns and these allow for bacteria growth. H_2SO_4 is one form of strong poison, because the poison's symptoms appear after five minutes from application on skin. If possible, treatment by Na_2CO_3 as antidote for H_2SO_4 , but the necrosis caused by bacterial infection should be treated use drugs. The main constituents of *Aloe Vera* gel are mucopolysaccharides (glucmannans, polymannoses, about 10% of total solids), enzymes, anthranoids, lignin, saponins, vitamins, amino acids (almost 50% of the total amount consisting of 8 of the 10 essential amino acids) and minerals (quantities not given). Total solids are in the range of 1.3 to 2%, the rest being water (Vinson *et al.*, 2005). *Aloe Vera* gel is obtained either from hand-filleted leaves of *Aloe barbadensis* or, by cold processing of the whole leaf, in which case the product usually also contains appreciable quantities of the latex material and anthranoids. The anthranoids in whole leaf extracts of *Aloe Vera* can however, be reduced to levels below 10mg/kg in the product (Reynolds and Dweck, 1999; Lee *et al.*, 2000; Hu *et al.*, 2003). Oliver (Oliver, 2012) indicates that *Aloe Vera* gel is used in veterinary medicine topically to promote wound healing on general skin wounds in all animals. It has also been recommended as a teat-dip in lactating cows, by intra mammary administration for (adjuvant) treatment of mastitis or high somatic cell counts, and by oral route in all food producing species as adjuvant treatment for a number of afflictions (ranging from anemia to infertility, mastitis and shock (Hu *et al.*, 2003; Oliver, 2012). Medicinal plants according to the World Health Organization (WHO) defines them as herbal preparations made by introducing plant materials to extraction, fractionation, purification, concentration, or other physical or biological processes, which may be produced as a basis for herbal products or for immediate consumption. In human medicine *Aloe Vera* gel is used topically to promote wound healing. Oral use as a general tonic for a number of indications, where scientific proof is outstanding, has also been described. *Aloe Vera* gel is also widely used in cosmetics (Ramachandra and Rao, 2008; Subramanian *et al.*, 2006; Saravanan *et al.*, 2010; Kedarnath *et al.*, 2012). Moreover, *Aloe Vera* has ulcerogenic activity (Sai *et al.*, 2014).

Objectives

1. To test the antimicrobial effects of Aloe vera aqueous and alcohol and Antibiotic (control) leaf extracts on *Staphylococcus aureus*.
2. To determine the effect of Aloe vera leaf extract different concentration on *Staphylococcus aureus*

MATERIALS AND METHODS

***Staphylococcus aureus*:** It was obtained from the microbiological laboratory of the Department of Pathology Medical lab, Faculty of Medicine ,University of Gezira, Wad

medani, Sudan during the period from February, 2016 to March, 2017.

***Aloe vera* plant:** Were obtained from the University of Gezira fields during February 2016 to March, 2017.

Preparation of Nutrient agar: This was a general-purpose cultured medium for bacteria. It was obtained in a dehydrated form. The constituent of the medium were beef extract, yeast extract, peptone, sodium chloride and agar. It was prepared according to the manufactures instruction by suspending 28g in one liter distilled water. The medium was allowed to boil until it was completely dissolved. The pH of medium was adjusted to pH 7.4 \pm 0.2 and then the medium was sterilized in an autoclave at 121°C (115b/in²) for 15 min (Harrigan, 1998).

Preparation of the crude extracts: *Aloe vera* leaf extract to prepare crude extract of fresh *Aloe vera* whole leaves were washed with distilled water, chopped into small pieces, air-dried and ground into powder. The *Aloe vera* mixed with 80% concentration of ethanol The pulp ethanol mix was then centrifuged at 3000rpm for 10 minutes and the supernatant collected was allowed to evaporate over a dry oven. The gelatinous extract thus prepared was weighed using distilled water, serial dilutions of 25g/75ml, 50g/50ml and 100mg (w/v) were made in order to obtain 25%, 50% and 100% concentrations, respectively.

Preparation of test organism: The nutrient agar were mixed well and poured on the sterile petri plates. The agar media on petri plates were allowed to set for few minutes. nutrient agar plates were inoculated with respective bacteria (*S.aureus*), and then incubated at 37° C for overnight. Each time, a fresh bacterial culture was prepared.

Antimicrobial agent: The antibacterial agent gentamicin was dissolved in distilled water. Further dilutions were made using the same solvent according to CLSI document M100-S18. Gentamicin was used in the concentrations 25%, 50% and 100%.

Antibacterial activity

Antibacterial activity was measured using paper disc diffusion method (Method of Saba *et al.*, 2011) was followed: The following steps were involved in paper disc diffusion method. The normal agar were mixed well and poured on the sterile petri plates. The agar media on petri plates were allowed to set harden for few minutes. nutrient agar plates were inoculated with respective bacteria. The small autoclaved discs of Whatmann filter paper were used. The test organism was spread on the petri plates by using sterilized glass spreader. During paper-disc diffusion method, the sterile discs were dipped in the different crude extracts of medicinal plants and antibiotic drugs with the help of sterilized forceps and placed on the Petri plates. Distilled water was used as a control to check the comparison of antibacterial activity with different crude extracts of medicinal plants. The petri plates were sealed with para film. Then, the petri plates were left at room temperature for 30 minute, to allow the diffusion of the test sample and then incubated at 37° C for overnight. The diameter of the zones of inhibition were measured in cm.

Statistical analysis: The obtained data was statistically analyzed by computer software MSTATC according to

analysis of variance (ANOVA); Duncan's Multiple Range Test was used for mean separation.

RESULTS

Two days post inoculation: The results depicted in Table (1) indicate that the high concentrations of Aloe vera aqueous extract (100%) used in this study had the highest inhibitory effects (18.2 mm) against the tested *Staphylococcus aureus*. However, this extract showed inhibition action of 12 mm even at minimal concentration (25%) used. The other concentrations of the aqueous phase (50 %) gave an inhibition zones of 15 mm. Control (Gentomycin) extract of Aloe vera at all concentrations shows an inhibitory effect against the *Staphylococcus aureus*. The highest inhibition zone obtained was 16.5 mm with the concentration of 100 % Concentrations of 25 and 50 % showed an inhibition zones of 7.75 and 6 mm, respectively.

Table 1. Effect of different Concentration of aqueous and alcoholic extracts of Aloe vera and Antibiotic on inhibition (mm) of *Staphylococcus aureus* using disc method at two days post inoculation

Treatments	Concentration %	Inhibition zones(mm)			Mean
		R1	R2	R3	
Ethanol	25	7.5	6	5.8	5.9
	50	10	9	9.8	9.4
	100	16	14	15	14.5
Aqueous	25	10	13	12	12
	50	15.8	16	14	15
	100	19.2	18.4	18	18.2
Gentamycin	25	6.5	8.5	7	7.75
	50	9	7	5	6
	100	17	15	18	16.5

Three days post inoculation: The results depicted in Table (2) indicate that the high concentrations of Aloe vera aqueous extract (100%) used in this study had the highest inhibitory effects (16.65 mm) against the tested *Staphylococcus aureus*. However, this extract showed inhibition action of 10.5 mm even at minimal concentration (25%) used in this study. The other concentrations of the aqueous phase (50 %) gave an inhibition zones of 13 mm. Control (gentamycin) extract of Aloe vera at all concentrations shows an inhibitory effect against the *Staphylococcus aureus*. The highest inhibition zone obtained was 16 mm with the concentration of 100 % Concentrations of 25 and 50 % showed an inhibition zones of 5.95 and 5 mm, respectively.

Table 2. Effect of different Concentration of aqueous and alcoholic extracts of Aloe vera and Antibiotic on inhibition (mm) of *Staphylococcus aureus* using disc method at three days post inoculation

Treatments	Concentration %	Inhibition zones(mm)			Mean
		R1	R2	R3	
Aqueous	25	10	11	10	10.5
	50	14	14	12	13
	100	18	16.4	16.9	16.65
Gentamycin	25	5	7.3	6.6	5.95
	50	8	6	5	5
	100	16	15	17	16

Four days post inoculation: The results depicted in (Table 3) indicate that the high concentrations of Aloe vera aqueous extract (100%) used in this study had the highest inhibitory

effects (16 mm) against the tested microorganisms. However, this extract showed inhibition action of 9 mm even at minimal concentration (25%) used in this study. The other concentrations of the aqueous phase (50 %) gave an inhibition zones of 11.5 mm. Control (Gentomycin) extract of Aloe vera at all concentrations shows an inhibitory effect against the *Staphylococcus aureus*. The highest inhibition zone obtained was 14 mm with the concentration of 100 % Concentrations of 25 and 50 % showed an inhibition zones of 4.5 and 4.5 mm, respectively.

Table 3. Effect of different Concentration of aqueous and alcoholic extracts of Aloe vera and Antibiotic on inhibition (mm) of *Staphylococcus aureus* using disc method at four days post inoculation

Treatments	Concentration%	Inhibition zones(mm)			Mean
		R1	R2	R3	
Aqueous	25	9	10	8	9
	50	12.8	12	11	11.5
	100	17	16	16	16
Gentamycin	25	3.5	5	4	4.5
	50	7	5	4	4.5
	100	15	13	15	14

DISCUSSION

This study showed that aqueous extract phase of Aloe vera gave better results compared to the antibiotic phase of the same extract at this study at all concentration tested. The broad antimicrobial action of the aqueous extract of the Aloe vera used in the study could be ascribed to the water soluble components which are naturally occurring in the plant materials. Other workers have reported There was no antimicrobial activity was reported using aqueous extract of A. vera leaves (Martinez et al., 1996). The antimicrobial activity of the extracts and their potency was quantitatively assessed by the presence or absence of inhibition zone and zone diameter. Only alcoholic extract was found to be a better solvent for extraction of antimicrobially active substances compared to water and hexane (Ahmad et al., 1998). (Subramanian et al., 2006). In other studies, the most effective antibiotic for gram positive is vancomycin than Gentamycin (Hoeger., 2004).

Aloe vera is a potent antimicrobial agent compared with the conventional antibiotics. The results of the study by Coopoosamy and Magwa., (2007) also revealed that lowest concentrations of ethyl acetate and ethanol crude extracts of Aloe excels resulted in complete inhibition of visible growth of pathogenic bacteria compared with the control antibiotics, chloramphenicol and streptomycin sulfate. The results of this study disagree with earlier studies that showed that the antibacterial activity of some Iranian medicinal plants were more significant in the solvent extracts compared with aqueous extracts in all the plants, indicating that the active principle(s) responsible for antibacterial activity were more soluble in organic solvents (Babu et al., 2007). Similarly, the solvent extract of cloves flower was also found to exhibit a significantly higher inhibitory effect on the caries-inducing properties of *Streptococcus mutans* compared with the crude aqueous extracts (Abd Rahim and Gulam.,2006). In other studies the methanolic and petroleum spirit extracts of Pelargonium essential oils were more potent antibacterial agents than the steam distilled volatile samples (Lis- Balchin et al.,1998).

Conclusion

It was observed that the highest concentration of the aqueous and ethanolic extract of the plant has significant effect on the bacteria isolates. They had lighter zone of inhibition compared to the antibacterial agents used as control. Similar result was reported by Adeleke *et al.* (2006).

REFERENCES

- Agarry OO, Olaleye MT. 2005. Bello-Micheal CO., Comparative activities of Aloe vera gel and leaf. *African Journal of Biotechnology*; 4(12):1413-1414.
- Ahlatat KS & Khatkar BS. 2011. Processing, food applications and safety of aloe vera products: a review. *J Food Sci Technol*, 48(5):525–33.
- Ahmad, J., Mehmood, Z. and Mohammad, F., 1998. Screening of some Indian medicinal plants for their antimicrobial properties. *Journal of Ethnopharmacology*. 62: 183-193.
- Alian, F, Rahimi., E, Shakerian, A, Momataz, H, Riah, M and Momeni, M. 2012. Antimicrobial resistance of staphylococcus aureus isolated from bovine, sheep and goat raw milk. *Global veterinaría* 8: 111 -114.
- Aloe ferox MILLER and its hybrids). London, UK: European Medicines Agency.
- Anderson, L.A., Phillipson, J.D. 1996. Herbal medicines, A Guide for Healthcare professionals, Pharmaceutical press, London.
- Annual report of the Oz FoodNet Network, 2010. Communicable Diseases Intelligence 36(3): E213–E241.
- Argudin MA, Mendoza MC, Rodicio MR. 2010. Food poisoning and Staphylococcus aureus enterotoxins. *Toxins* 2(7):1751–1773.
- Asao T, Kumeda Y, Kawai T, Shibata T, Oda H, Haruki K, Nakazawa H, Kozaki S. 2003. An extensive outbreak of staphylococcal food poisoning due to low-fat milk in Japan: Estimation of enterotoxin A in the incriminated milk and powdered skim milk. *Epidemiology and Infection* 130:33–40.
- Aydin A, Sudagidan M, Muratoglu K. 2011. Prevalence of staphylococcal enterotoxins, toxin genes and genetic relatedness of foodborne Staphylococcus aureus strains isolated in the Marmara region of Turkey. *International Journal of Food Microbiology* 148:99–106.
- Boudreau MD, Beland FA, Nichols JA, Pogribna M. 2013. Toxicology and carcinogenesis studies of a noncolorized whole leaf extract of Aloe barbadensis Miller (Aloe vera) in F344/N rats and B6C3F1 mice (drinking water study). *Natl Toxicol Program Tech Rep Ser*, 577(577):1–266. PMID:24042237.
- CDC, 2012. Summary of notifiable diseases - United States, 2010. *Morbidity and Mortality Weekly Report* 59(53):1–111.
- Channe Gowda D, Neelisiddaiah B, Anjaneyalu YV 1979. Structural studies of polysaccharides from Aloe vera. *Carbohydr Res*, 72:201–5.
- Committee of Experts on Cosmetic Products 2008. Aloe extracts with anthraquinones. Active ingredients used in cosmetics: safety survey. Strasbourg, France: Council of Europe Publishing; pp. 9–27.
- Coopoosamy RM, Magwa ML., Traditional use, antibacterial activity and antifungal activity of crude extract of Aloe excelsa. *African Journal of Biotechnology* (2007); (20):240-2410.
- Cosmetic Ingredient Review Expert Panel 2007. Final report on the safety assessment of AloeAndongensis Extract, Aloe Andongensis Leaf Juice, Aloe Arborescens Leaf Extract, Aloe Arborescens Leaf Juice, Aloe Arborescens Leaf Protoplasts, Aloe Barbadosensis Flower Extract, Aloe Barbadosensis Leaf, Aloe Barbadosensis Leaf Extract, Aloe Barbadosensis Leaf Juice, Aloe Barbadosensis Leaf Polysaccharides, Aloe Barbadosensis Leaf Water, Aloe Ferox Leaf Extract, Aloe Ferox Leaf Juice, and Aloe Ferox Leaf Juice Extract. *Int J Toxicol*, 26(1):Suppl 2: 1–50.
- Crossley KB and Archer GL, The Staphylococci in Human Disease., Churchill Livingstone, 1997.
- Dal’Belo SE, Gaspar LR, Maia Campos PM. 2006. Moisturizing effect of cosmetic formulations containing Aloe vera extract in different concentrations assessed by skin bioengineering techniques. *Skin Res Technol*, 12(4):241–6.
- Davidson PM, Taylor TM. 2007. Chemical preservatives and natural antimicrobial compounds. Ch 33 In: Doyle MP, Beuchat LR(eds) *Food microbiology: Fundamentals and frontiers*. 3rd ed, ASM Press, Washington D.C., p. 713–745.
- Dentali S. 2013. “Nondecolorized” essential qualifier for NTP aloe vera study material. *Toxicol Sci*, 133(2):342.
- EFSA 2013. The European Union summary report on trends and sources of zoonoses, zoonotic agents and foodborne outbreaks in 2011. *EFSA Journal* 11(4):3129.
- EFSA. 2012. The European Union summary report on trends and sources of zoonoses, zoonotic agents and foodborne outbreaks in 2010. *EFSA Journal* 10(3):2597.
- EI-Jakee, S, A. Marouf, Nagwa S. Ata, Eman H. Abdel-Rahman, , Sherein I. Abd El-Moez, A.A. Samy and 2Walaa E. El-Sayed ,2014. Rapid Method for Detection of Staphylococcus aureus Enterotoxins in Food, *Global Veterinaría* 11 : 335-341.
- Elsohly MA, Gul W, Avula B, Khan IA. 2007. Determination of the anthraquinones aloe-emodin and aloin-A by liquid chromatography with mass spectrometric and diode array detection. *J AOAC Int*, 90(1):28–42. PMID:17373434.
- EMA, 2006. Community herbal monograph on Aloe barbadensis MILLER and on Aloe (various species, mainly EMA, 2006. Community herbal monograph on Aloe barbadensis MILLER and on Aloe (various species, mainly Aloe ferox MILLER and its hybrids). London, UK: European Medicines Agency.
- Eur Ph; European Pharmacopoeia 2008. European pharmacopoeia. 7.0. Strasbourg, France: European Directorate for the Quality of Medicines & HealthCare.
- Evenson ML, Hinds MW, Berstein RS, Bergdoll MS. 1988. Estimation of human dose of staphylococcal enterotoxin A from a large outbreak of staphylococcal food poisoning involving chocolate milk. *International Journal of Food Microbiology* 7:311–316.
- FDA 2012. Bad bug book: Foodborne pathogenic microorganisms and natural toxins handbook, 2nd ed. US Food and Drug Administration, Silver Spring, p. 8792. <http://www.fda.gov/Food/FoodborneIllness-Contaminants/CausesOfIllnessBadBugBook/ucm2006773.htm>. Accessed 27 March (2013).
- Femenia A, Sanchez E, Simal S, Rosselló C. 1999. Compositional features of polysaccharides from Aloe vera (Aloe barbadensis Miller) plant tissues. *Carbohydr Polym*, 39(2):109–17. doi:10.1016/S0144- 8617(98)00163-5.

- Figuroa G, Navarrete P, Caro M, Troncoso M, Faundez G 2002. Carriage of enterotoxigenic *Staphylococcus aureus* in food handlers. *Revista Medica De Chile* 130(8):859–864.
- Fitzgerald, J.R., Sturdevant, D.E., Mackie, S.M., Gill and S.R., Musser, J.M., 2001. Evolutionary genomics of *Staphylococcus aureus*: insights into the origin of methicillin-resistant strains and the toxic shock syndrome epidemic. *Proceeding of the Natural Academic Science*, 98: 8821-8826.
- Foster T, *Staphylococcus*, in: Baron (Ed.), *Medical Microbiology*, University of Texas Medical Branch at Galveston, Texas, (1996).
- Gavimath, C.C., Ramachandra, Y.L., Padmalatha Rai, S., Sudeep, H.V., Sujan, P.S., Ganapathy and Kavitha, B.T. (2008). Antibacterial activity of *Aegle marmelos* Correa leaves extract, *Asian Journal of Bioscience*. 3(2): 333-336.
- Gaze JE. 1985. The effect of oil on the heat resistance of *Staphylococcus aureus*. *Food Microbiology* 2:277–283.
- Grindlay D & Reynolds T. 1986. The Aloe vera phenomenon: a review of the properties and modern uses of the leaf parenchyma gel. *J Ethnopharmacol*, 16(2-3):117–51. doi:10.1016/0378-8741(86)90085-1 PMID:3528673.
- Groom QJ & Reynolds T. 1987. Barbaloin in aloe species. *Planta Med*, 53(4):345–8.
- Hackbarth, C.J., and Chambers, H.F., 1989. Methicillin-resistant staphylococci: detection methods and treatment of infections. *Antimicrobial Agents Chemother*. 33:995 – 999.
- Hall G, Kirk MD, Becker N, Gregory JE, Unicomb L, Millard G, Stafford R, Lalor K. 2005. Estimating foodborne gastroenteritis, Australia. *Emerging Infectious Diseases* 11(8):1257–1264.
- Hamman JH. 2008. Composition and applications of Aloe vera leaf gel. *Molecules*, 13(8):1599–616. doi:10.3390/molecules13081599 PMID:18794775.
- Harrigan, W.F. 1998. *Laboratory Methods in Food Microbiology*. Academic Press, San Diego.
- Hatakka M, Bjorkroth KJ, Asplund K, Maki-Petays N, Korkeala HJ. 2000. Genotypes and enterotoxigenicity of
- Howard BJ and Kloos WE, *Staphylococci*, in: Howard BJ, Klass J, J RS, Weissfeld AS, and Tilton RC (Eds.), *Clinical and Pathogenic Microbiology*, Mosby, Washington, 1987, pp. 231-234.
- Hu, Y., Xu J and Q. Hu, 2003. Evaluation of antioxidant potential of Aloe vera (*Aloe barbadensis* Miller) extracts. *Journal Agriculture. Food Chemistry*, 51: 7788-7791.
- ICMSF, 1996. *Staphylococcus aureus*. Ch 17 In: *Microorganisms in food 5: Microbiological specifications of food pathogens*. Blackie Academic and Professional, London, p. 299–333.
- Jehan Bakht, Amjad Islam and Mohammed Shafi. 2011. antimicrobial potentials of *eclipta alba* by well diffusion method., *Pak. J.Bot.*, 43: 169-174.
- Joseph B & Raj SJ 2010. Pharmacognostic and phytochemical properties of Aloe vera Linn—an overview. *International Journal of Pharmaceutical Sciences Review and Research*, 4:106–110.
- JP XVI, *The Japanese Pharmacopoeia* 2011. The Japanese Pharmacopoeia 16th ed. English Version, Tokyo, Japan: Ministry of Health, Labour and Welfare.
- Kaithwas G, Kumar A, Pandey H, 2008. Investigation of comparative antimicrobial activity of Aloe vera gel and juice. *Pharmacology online*; 1:239-243.
- Kedarnath, N.K., Surekh.a, Ramesh. S, Mahantesh. S. P and Patil C.S. 2012. Phytochemical screening and antimicrobial activity of Aloe vera L. *World Research Journal of Medicinal & Aromatic Plants* 1 : 11-1.
- Kennedy J, Blair IS, McDowell DA, Bolton DJ. 2005. An investigation of the thermal inactivation of *Staphylococcus aureus* and the potential for increased thermo tolerance as a result of chilled storage. *Journal of Applied Bacteriology* 99:1229–1235.
- Kitai S, Shimizu A, Kawano J, Sato E, Nakano C, Kitagawa H, Fujio K, Matsumura K, Yasuda R, Inamoto T 2005. Prevalence and characterization of *Staphylococcus aureus* and enterotoxigenic *Staphylococcus aureus* in retail raw chicken meat throughout Japan. *The Journal of Veterinary Medical Science* 67(3):269–274.
- Kloos WE and Bannerman TL. Update on clinical significance of coagulase-negative staphylococci. *Clinical Microbiology Reviews* 7[1], 117-140. 1994.
- Kloos WE and Lambe DWJ, *Staphylococcus*, in: Barlows A, Hausler WJ, Hermann KL, Isenberg HD, and Shadomy HJ (Eds.), *Manual of Clinical Microbiology*, ASM, Washington D.C, 1991, pp. 222-237.
- Kluytmans J, van Belkum A, and Verbrugh H. Nasal carriage of *Staphylococcus aureus*: epidemiology, underlying mechanisms, and associated risks. *Clinical Microbiology Reviews* 10[3], 505-520. (1997).
- Lachenmeier K, Kuepper U, Musshoff Fet al. 2005. Quality control of Aloe vera beverages. *Electronic Journal of Environmental, Agricultural and Food Chemistry*, 4:1033–1042.
- Lee, K., S. Weintraub and B. Yu, 2000. Isolation and identification of a phenolic antioxidant from Aloe barbadensis. *Free Radical Biology and Medicine Journal*, 28: 261-265.
- Leon L. 2003. The medicinal plant Aloe. *Ganzheitliche Tiermedizin*, 17:138–143.
- Lim E, Lopez L, Borman A, Cressey P, Pirie R. 2012. Annual report concerning foodborne disease in New Zealand 2011. Ministry for Primary Industry, New Zealand. <http://www.foodsafety.govt.nz/science-risk/human-health-surveillance/foodborne-diseaseannual-reports.htm>. Accessed 11 April 2013.
- Liu GY, Essex A, Buchanan JT, Datta V, Hoffman HM, Bastian JF, Fierer J, and Nizet V. *Staphylococcus aureus* golden pigment impairs neutrophil killing and promotes virulence through its antioxidant activity.
- Mandal G & Das A 1980. Structure of the D-galactan isolated from Aloe barbadensis Miller. *Carbohydr Res*, 86(2):247–57.
- Mangena T. 1999. Comparative evaluation of the antimicrobial activities of essential oils of *Artemisia afra*, *Pteronia incana* and *Rosmarinus officinalis* on selected bacteria and yeast strains. *Letent. Applied Microbiology*; 28(4):291-296.
- Montville TJ, Matthews KR 2008. *Food microbiology: An introduction*. 2nd ed, ASM Press,
- Nema V, Agrawal R, Kamboj DV, Goel AK, Singh L. 2007. Isolation and characterization of heat resistant Enterotoxigenic *Staphylococcus aureus* from a food poisoning outbreak in Indian subcontinent. *International Journal of Food Microbiology* 117:29–35.
- Newton LE. 2004. Aloes in habitat. In: Reynolds T, editor. *Aloes: the genus Aloe*. Boca Raton (FL), USA: CRC Press; pp. 3–14.
- Ni Y, Turner D, Yates KM, Tizard I. 2004. Isolation and characterization of structural components of Aloe vera L. leaf pulp. *Int Immunopharmacol*, 4(14):1745–55.

- NLM. 2012. Products that contain active ingredient - Aloe vera. Dietary supplements labels database. United
- Normanno G, La Salandra G, Dambrosio A, Quaglia NC, Corrente M, Parisi A, Santagada G, Firinu A, Crisetti E, Celano GV. 2007. Occurrence, characterization and antimicrobial resistance of enterotoxigenic Staphylococcus aureus isolated from meat and dairy products. *International Journal of Food Microbiology* 115:290–296.
- O’Neil MJ, Heckelman PE, Koch CB et al. 2006. The Merck Index - An Encyclopedia of Chemicals, Drugs, and Biologicals. 14th Ed. Version 14.6. Whitehouse Station (NJ), USA: Merck & Co., Inc.
- Oliver, G. 2012. Aloe Vera Gel Research Review An overview of its clinical uses and proposed mechanisms of action. *Natural Medical Journal* 4: 120-133.
- OzFoodNet (2012b) Oz FoodNet Quarterly report, 1 July to 30 September 2011. Communicable Diseases Intelligence 36(2):E188–E195.
- OzFoodNet 2012. Monitoring the incidence and causes of diseases potentially transmitted by food in Australia:
- Park YI, Jo TH 2006. Perspectives of industrial application of Aloe vera. In: Park YI, Lee SK, editors. *New Perspectives on Aloe*. New York (NY), USA: Springer Science+Business Media; pp. 191–200.
- Peacock SJ, de Silva I, and Lowy FD. What determines nasal carriage of Staphylococcus aureus? *Trends in microbiology* 9[12], 605-610. (2001).
- Pelley RP, Martini WJ, Liu DQ et al. 1998. Multiparameter analysis of commercial “Aloe vera” materials and comparison to Aloe barbadensis Miller extracts. *Subtropical Plant Science*, 50:1–14.
- Pellizzoni M, Molinari GP, Lucini L. 2011. Stability of the main Aloe fractions and Aloe-based commercial products under different storage conditions *Agrochimica*, 5:288–296.
- Pinchuk IV, Beswick EJ, Reyes VE. 2010. Staphylococcal enterotoxins. *Toxins* 2:2177–2197.
- Plata K, Rosato AE, and Wegrzyn G. Staphylococcus aureus as an infectious agent: overview of biochemistry and molecular genetics of its pathogenicity. *Acta Biochimica Polonica* 56[4], 597-612. (2009).
- Raj HD, Bergdoll MS. 1969. Effect of enterotoxin B on human volunteers. *Journal of Bacteriology* 98(2):833–834.
- Ramachandra, C and Rao.P, 2008. Processing of Aloe vera leaf gel: a review. *American journal of Agricultural and Biological Sciences*, 3:502-510.
- Ray C and Ryan KJ, Sherris *Medical Microbiology: An Introduction to Infectious Diseases.*, 2003.
- Reynolds T. 2004. Aloe chemistry. In: Reynolds T, editor. *Aloes: The genus Aloe*. 38th Ed. Boca Raton (FL), USA: CRC Press; pp. 39–74.
- Reynolds, T and A. Dweck, 1999, Aloe Vera leaf gel: a review update. *Journal Ethnopharmacol*, 68: 3-37.
- Rodríguez ER, Martín JD, Romero CD. 2010. Aloe vera as a functional ingredient in foods. *Crit Rev Food Sci Nutr*, 50(4):305–26.
- Rodríguez ER, Martín JD, Romero CD. 2010. Aloe vera as a functional ingredient in foods. *Crit Rev Food Sci Nutr*, 50(4):305–26.
- Saba Irshad, Muneeba Butt and Hira Younus, 2011. “In-vitro antibacterial of Aloe barbadensis Miller (Aloe vera)”, *International Research Journal of pharmaceuticals* Vol.01, Issue 02, pp. 59-64.
- Saccù D, Bogoni P, Procida G. 2001. Aloe exudate: characterization by reversed phase HPLC and headspace GC-MS. *J Agric Food Chem*, 49(10):4526–30.
- Saravanan, P., Ramya .V, Sridhar, H, Balamurugan .V and Umamaheswari S, 2010. Antibacterial activity of Allium sativum L. on pathogenic bacterial strains. *Global Veterinaria* 4 :519-522.
- Sehgal I, Winters WD, Scott M, Kousoulas K. 2013. An in vitro and in vivo toxicologic evaluation of a stabilized aloe vera gel supplement drink in mice. *Food Chem Toxicol*, 55:363–70.
- Seo KS, Bohach GA. 2007. Staphylococcus aureus. Ch 22 In: Doyle MP, Beuchat LR (eds) *Food microbiology: Fundamentals and frontiers*. 3rd ed, ASM Press, Washington D.C., p. 493–518.
- Shao A, Broadmeadow A, Goddard G, Bejar E, Frankos V 2013. Safety of purified decolorized (low anthraquinone) whole leaf Aloe vera (L) Burm. f. juice in a 3-month drinking water toxicity study in F344 rats. *Food Chem Toxicol*, 57:21–31.
- Simon SS, Sanjeev S. 2007. Prevalence of enterotoxigenic Staphylococcus aureus in fishery products and fish processing factory workers. *Food Control* 18(12):1565–1568.
- Staphylococcus aureus isolated from the hands and nasal cavities of flight catering employees. *Journal of Food Protection* 63(11):1487–1491.
- States National Library of Medicine. Available from: <http://www.dslid.nlm.nih.gov/dslid/>; accessed 5 June 2014.
- Steenkamp V & Stewart MJ 2007. Medicinal applications and toxicological activities of Aloe products. *Pharmaceutical Biology*, 45(5):411–20.
- Steenkamp V & Stewart MJ. 2007. Medicinal applications and toxicological activities of Aloe products. *Pharmaceutical Biology*, 45(5):411–20. doi:10.1080/13880200701215307.
- Stewart CM. 2003. Staphylococcus aureus and staphylococcal enterotoxins. Ch 12 In: Hocking AD (ed) *Foodborne microorganisms of public health significance*. 6th ed, *Australian Institute of Food Science and Technology* (NSW Branch), Sydney, p. 359–380.
- Subramanian S, Kumar DS, Arulselvan P, Senthikumar GP., In vitro antibacterial and antifungal activities of ethanolic extract of Aloe vera leaf gel. *Journal of Plant Science* (2006); 1(4):348-355.
- Subramanian, S., D. Sathish. Kumar and P. Arulselvan, 2006. Wound healing potential of Aloe vera leaf gel studied in experimental rabbits. *Asian Journal Biochemistry*, 1: 178-185.
- T. The Journal of Experimental Medicine 202[2], 209-215. (2005)
- Talarico F, Rocchia E, Nero Id. 1997. Prevalence of enterotoxigenic Staphylococcus in foodhandlers in the province of Catanzaro (Italy). *Igiene Moderna* 107(2):137–142.
- Ulbricht C, Armstrong J, Basch E, Basch S, Bent S, Dacey Cet al. 2007. An evidence-based systematic review of Aloe vera by the natural standard research collaboration. *J Herb Pharmacother*, 7(3-4):279–323.
- Vinson, J., H. Al kharrat and L. Andreoll, 2005. Effect of Aloe Vera preparations on the human bioavailability of vitamins C and E. *Phytomedicine Journal*, 12: 760-765.
- Washington D.C. NCCAM (2012). Aloe vera. Bethesda (MD): National Center for Complementary and Alternative Medicine. Available from: <http://nccam.nih.gov/health/aloevera>, accessed 5 June 2014.

- Wertheim HFL, Melles DC, Vos MC, van Leeuwen W, van Belkum A, Verbrugh HA, and Nouwen JL. The role of nasal carriage in *Staphylococcus aureus* infections. *The Lancet Infectious Diseases* 5[12], 751- 762. 2005.
- WHO (1999). Aloe and Aloe vera gel. WHO Monographs on selected medicinal plants. Vol. 1. Geneva, Switzerland: World Health Organization; pp. 33–49(available from <http://apps.who.int/medicinedocs/en/d/Js2200e/5.html>).
- Wilkinson BJ, Biology, in: Crossley KB and Archer GL (Eds.), *The Staphylococci in Human Diseases*. Churchill Livingstone, London, 1997. pp. 1-38.
- Yaron A. 1993. Characterization of Aloe vera gel before and after autodegradation, and stabilization of the natural fresh gel. *Phytother Res*, 7(7):11.
